

Article

Characterization of Stem-like Circulating Tumor Cells in Pancreatic Cancer

Lei Zhu ¹, Barbara Hissa ¹, Balázs Gyórfy ^{2,3}, Johann-Christoph Jann ⁴, Cui Yang ¹, Christoph Reissfelder ^{1,5} and Sebastian Schölch ^{1,5,*}

¹ Department of Surgery, Universitätsmedizin Mannheim, Medical Faculty Mannheim, Heidelberg University, 68167 Mannheim, Germany; Lei.Zhu@medma.uni-heidelberg.de (L.Z.); Barbara.Hissa@medma.uni-heidelberg.de (B.H.); Cui.Yang@umm.de (C.Y.); christoph.reissfelder@umm.de (C.R.)

² 2nd Department of Pediatrics, Semmelweis University, H-1094 Budapest, Hungary; gyorffy.balazs@yahoo.com

³ TTK Cancer Biomarker Research Group, Institute of Enzymology, H-1117 Budapest, Hungary

⁴ Department of Medicine III, Universitätsmedizin Mannheim, Medical Faculty Mannheim, Heidelberg University, 68167 Mannheim, Germany; Johann-Christoph.Jann@medma.uni-heidelberg.de

⁵ German Cancer Consortium (DKTK) & German Cancer Research Center (DKFZ), 69120 Heidelberg, Germany

* Correspondence: sebastian.schoelch@medma.uni-heidelberg.de; Tel.: +49-621-383-5152

Received: 17 February 2020; Accepted: 13 May 2020; Published: 15 May 2020

Abstract: Pancreatic ductal adenocarcinoma (PDAC) is the fourth most frequent cause of death from cancer. Circulating tumor cells (CTCs) with stem-like characteristics lead to distant metastases and thus contribute to the dismal prognosis of PDAC. Our purpose is to investigate the role of stemness in CTCs derived from a genetically engineered mouse model of PDAC and to further explore the potential molecular mechanisms. The publically available RNA sequencing dataset GSE51372 was analyzed, and CTCs with (CTC-S) or without (CTC-N) stem-like features were discriminated based on a principal component analysis (PCA). Differentially expressed genes, weighted gene co-expression network analysis (WGCNA), and further functional enrichment analyses were performed. The prognostic role of the candidate gene (*CTNNB1*) was assessed in a clinical PDAC patient cohort. Overexpression of the pluripotency marker *Klf4* (Krüppel-like factor 4) in CTC-S cells positively correlates with *Cttnb1* (β -Catenin) expression, and their interaction presumably happens via protein–protein binding in the nucleus. As a result, the adherens junction pathway is significantly enriched in CTC-S. Furthermore, the overexpression of *Cttnb1* is a negative prognostic factor for progression-free survival (PFS) and relapse-free survival (RFS) in human PDAC cohort. Overexpression of *Cttnb1* may thus promote the metastatic capabilities of CTCs with stem-like properties via adherens junctions in murine PDAC.

Keywords: pancreatic cancer; PDAC; circulating tumor cells; CTC; stem cells; stem-like; stemness; adherens junctions; epithelial-mesenchymal transition; EMT

1. Introduction

With 57,000 new cases and 46,000 deaths annually, pancreatic ductal adenocarcinoma (PDAC) is the 9th most frequent malignant disease in males and 8th most prevalent in females and the 4th most frequent cause of death in both genders [1]. At the time of diagnosis, only a fraction of patients are amenable to surgical resection of the tumor. However, even patients who undergo complete surgical resection are at a high risk of either local or systemic recurrence [2].

In order to study the molecular mechanisms of pathogenesis and metastasis of pancreatic cancer, several genetically induced mouse models (GEMMs) have been established in recent decades [3–6].

The most prominent and best-characterized model is the LSL-*Kras*^{G12D}, LSL-*Trp53*^{R172H}, *Pdx1-Cre* (KPC) mouse model [6].

It is now well established that circulating tumor cells (CTCs) are an integral part of the metastatic cascade in malignant disease [7–18]. CTCs are shed from the primary tumor, survive in circulation, and ultimately colonize distant organs where they establish clinically overt metastases [9,16]. However, while thousands or millions of CTCs are shed into the bloodstream over time, the number of metastases is several orders of magnitude lower [19]. Therefore, it can be assumed that only a small fraction of CTCs are actually tumorigenic and thus clinically relevant. In order to achieve the invasive phenotype required to leave the primary tumor bulk and enter circulation, CTCs from epithelial tumors undergo a process called epithelial–mesenchymal transition (EMT). During this process, the cells acquire mesenchymal properties (e.g., migratory capability) while downregulating epithelial traits [20,21]. This process is reverted during mesenchymal-epithelial transition (MET). As both EMT and MET dynamically fluctuate, CTCs in circulation exhibit a high degree of plasticity and represent a heterogeneous population consisting of epithelial, mesenchymal, and intermediary cells [20,22].

In a previous study, CTCs from the GEMM (LSL-*Kras*^{G12D}, LSL-*Trp53*^{flox/flox} or *flox/+*, *Pdx1-Cre*) were isolated and submitted to single-cell RNA sequencing. The raw data of this experiment was released to the Gene Expression Omnibus (GEO) database, and it is available for analysis [23]. In previous studies from our own group, we were able to demonstrate that, apart from immune evasive capabilities, CTCs also possess stem cell properties [13,17]. The main goal of this study was therefore to investigate the role of stem cell properties in murine CTCs of PDAC and to further characterize CTCs with stem cell properties.

2. Results

Details regarding the study workflow are depicted in Figure 1.

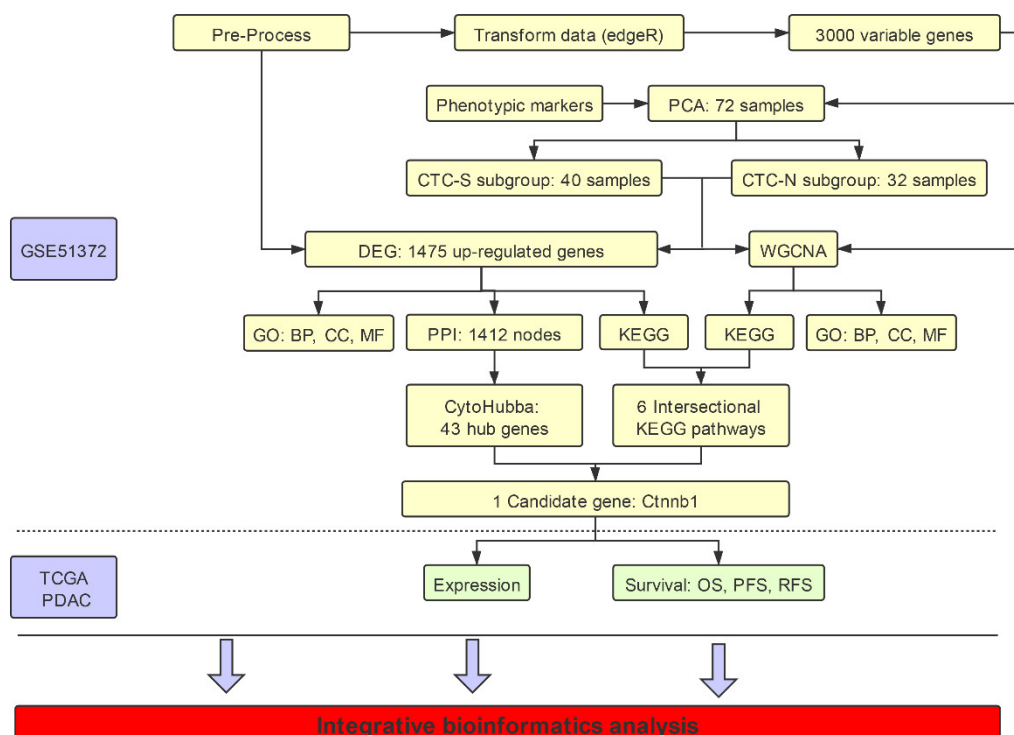


Figure 1. Illustration of the workflow in the integrative bioinformatics analysis. Data set GSE51372 from the Gene Expression Omnibus (GEO) database was used, which contains 75 murine pancreatic CTCs single-cell sequencing datasets. After filtering out three samples with low quality, we further excluded low expression genes, and EdgR transformed the remaining 72 samples. CTCs with stem-like features (CTC-S) and CTC with non-stem-like features (CTC-N) were defined based on the principal component analysis (PCA) of the top 3000 most variable genes. We performed weighted gene

co-expression network analysis (WGCNA) on these 3000 genes, while all read counts of 11,931 genes were used to analyze the differentially expressed genes. Further functional enrichment analysis, including the Kyoto Encyclopedia of Genes and Genomes (KEGG) pathways, Gene Ontology (GO) aspects on both differentially expressed genes and WGCNA results was performed. The cytoHubba plugin of Cytoscape 3.5.1 was employed to identify hub genes. Candidate pathways (adherens junction) and genes (*Ctmb1*) were identified. The clinical value of *KLF4* and *CTNNB1* was validated in the TCGA (The Cancer Genome Atlas) pancreatic adenocarcinoma (PDAC) cohort. The workflow was plotted using ProcessOn (available online: <https://www.processon.com/diagrams>, accessed on 23 March 2020).

2.1. Pancreatic CTCs Can Be Clustered into Stem-Like and Non-Stem like Categories

After pre-processing, we kept 72 samples for the following study, and 11,931 genes were obtained (Figure S1A,B). In order to classify CTCs into subgroups, the top 3000 genes that presented the highest variability in expression were chosen for the principal component analysis (PCA) [19,24,25]. From these 3000 genes, epithelial (*Epcam*, *Krt7*, *Krt8*, *Krt18*, and *Krt19*) [23], mesenchymal (*Fn1*, *S100a4* and *Vim*) [26], stemness/pluripotency markers (*Aldh1a1*, *Aldh1a2*, *Cd24a*, *Cd44*, and *Klf4*) [25,27,28] and the proliferation marker *Mki67* [29] were included in subsequent analyses.

All 72 individual CTCs could be divided into three clusters (Figure S2A, B). However, this study focuses on whether the CTCs have stem-like features or not, a dichotomous classification; therefore, the CTCs were divided into clusters based on stem-like features. According to the PCA loading plot (Figure 2A), the cluster located in the first quadrant (blue dots) exhibited stemness markers and was defined as CTC-S (CTCs with stem-like features) (Figure 2B). The other two clusters were merged (red dots) and named CTC-N (CTC without stemness features). The correlation heatmap with hierarchical clustering showed a similar result (Figure 2C).

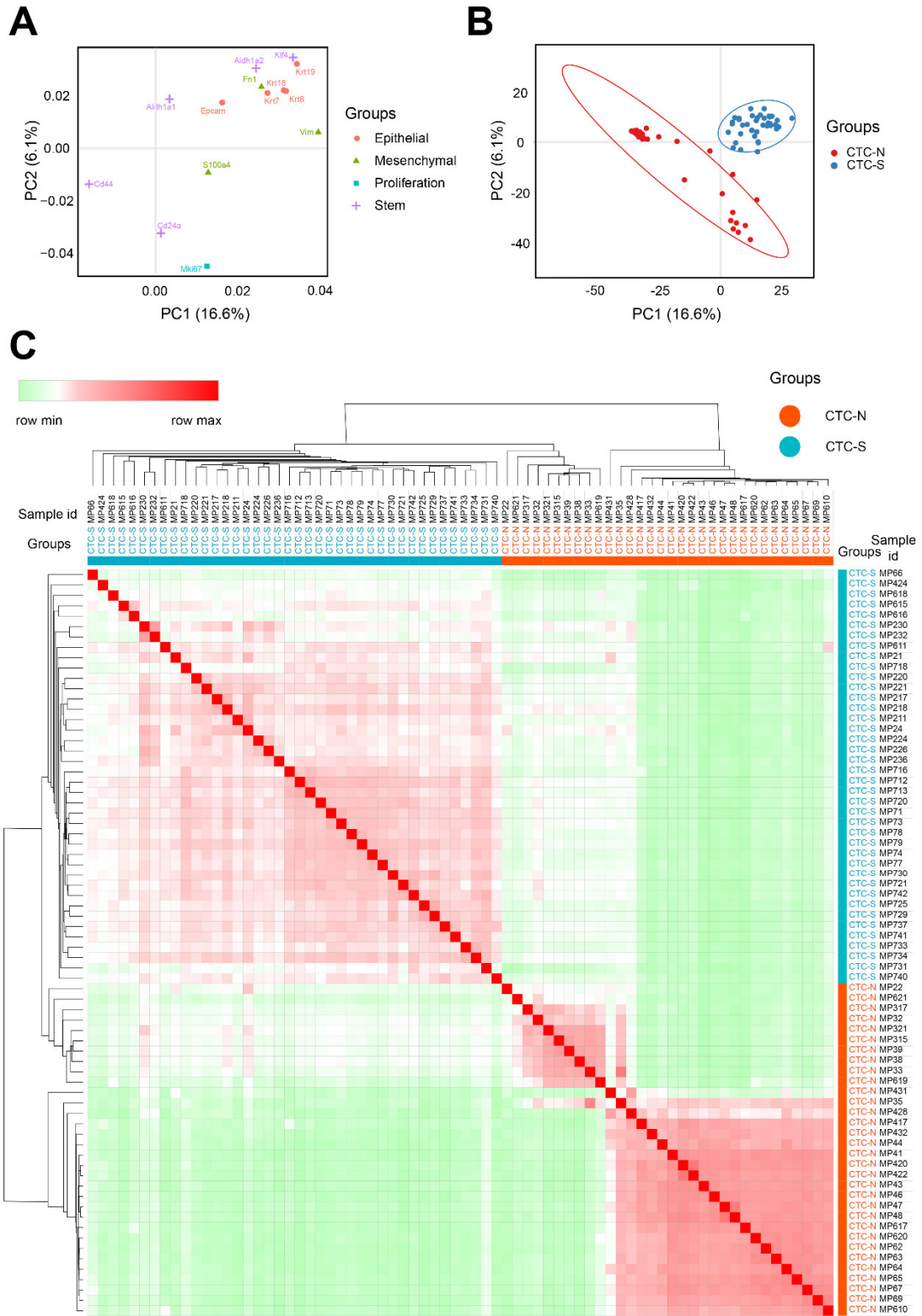


Figure 2. Defining the CTC-S and CTC-N subgroups. **(A)** Principal component analysis (PCA) loading plot of the marker set. The x and y axes represent the principal components 1 (PC1) = 16.6% variance and PC2 = 6.1% variance, respectively. 3 of the 5 stem markers (Aldh1a1, Aldh1a2, and Klf4) are located in the first quadrant, indicating that stem markers tend to correlate with both PC1 and PC2 positively. The PCA loading data were downloaded from the ClustVis and visualized by the imageGP (available online: <http://www.ehbio.com/ImageGP/index.php/Home/Index/index.html>, accessed on 23 April 2020). **(B)** PCA scores plot of 72 samples. All samples were divided into three clusters. The cluster located in the first quadrant was defined as CTCs with stem-like features (CTC-S) since they

present with stem markers PCA loadings in (A), as the other two clusters were combined and defined as CTC with non-stem-like features (CTC-N). The corresponding ellipses were plotted based on a 95% probability from the same group. (C) The correlation heatmap was visualized by MORPHEUS (available online: <https://software.broadinstitute.org/morpheus/>, accessed on 7 March 2020). We chose the average linkage method to perform the hierarchical clustering. The heatmap demonstrates the distinct subgroups of CTC-S and CTC-N. The colors of the square matrices illustrate the Pearson's correlation coefficient, with red indicating a strong correlation and green a weak correlation. All samples are listed in the same order in both horizontal and vertical axes.

2.2. Both Epithelial and Mesenchymal Markers are Expressed on CTC-S

To evaluate each cell within their respective subgroups in more detail, we expanded the marker pool, as not all markers of interest were listed in the 3000 most variable genes. EMT markers (*Snai1*, *Snai2*, *Twist1*, *Zeb1*, and *Zeb2*), epithelial markers (*Cdh1*, *Egfr*, *Epcam*, *Krt7*, *Krt8*, *Krt18*, and *Krt19*) [23], mesenchymal markers (*Cdh2*, *Fn1*, *Itga5*, *Sdc1*, *S100a4*, and *Vim*) [20,23], pancreatic cancer stemness markers (*Abcg2*, *Aldh1a1*, *Aldh1a2*, *Cd24a*, *Cd44*, *Cxcr4*, *Met*, *Prom1*, *Klf4*, *Nanog* and *Sox2*) [30,31], and a proliferation marker (*Mki67*) were included to plot the heatmap (Figure 3A).

A high degree of expression heterogeneity was observed for most markers. Compared with CTC-N, CTC-S cells demonstrated an increased expression of epithelial markers such as *Egfr*, *Epcam*, and *keratins*. Parallely, the mesenchymal markers *Fn1* [$\log_2(\text{fold change})$ (FC) = 4.07, false discovery rate (FDR) = 1.42×10^{-5}] and *Vim* ($\log_2\text{FC}$ = 6.37, FDR = 5.89×10^{-10}) were up-regulated in the CTC-S group, resulting in a phenotype with both epithelial and mesenchymal characteristics. Notably, stemness transcripts showed variable expression, while *Aldh1a2* ($\log_2\text{FC}$ = 4.98, FDR = 1.61×10^{-8}) and *Klf4* ($\log_2\text{FC}$ = 9.41, FDR = 1.45×10^{-21}) were enriched in CTC-S, *Cd44* was significantly down-regulated ($\log_2\text{FC}$ = -2.57, FDR = 0.012) (Figure 3A).

2.3. The Adherens Junction Pathway is Functionally Enriched in CTC-S

Compared with CTC-N, 1475 genes were significantly up- and 247 significantly down-regulated in the CTC-S group (Figure S3A, B).

We next constructed a protein–protein interaction (PPI) network of up-regulated genes by STRING (available online: <https://string-db.org/>, accessed on 19 August 2019) [32], establishing 1412 nodes and 6695 interactions (Figure not shown). We used the cytoHubba plugin in Cytoscape to narrow this number down to 43 genes, which were subsequently defined as hub genes.

The biological domains of biological process (BP), cellular component (CC), and molecular function (MF) were utilized to describe the biological domain of genes enriched in the CTC-S group. The cellular components' anchoring junction [Gene Ontology (GO): 0070161, fold enriched = 3.56, FDR = 1.55×10^{-40}] and adherens junction (GO: 0005912, fold enriched = 3.53, FDR = 9.73×10^{-39}) were significantly functionally enriched. The enrichment of cell adhesion molecule binding (GO: 0050839, fold enriched = 3.34, FDR = 6.48×10^{-31}) was observed in the MF domain (Figure 3B).

While GO enrichment analysis focuses on the functions of the genes, the Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway analysis mainly reflects the interaction network and the signaling pathways of the requested genes. In this analysis, several signaling pathways were significantly enriched, including adherens' junction (KEGG: 04520, fold enriched = 3.42, FDR = 1.47×10^{-5}) and fluid shear stress and atherosclerosis pathways (KEGG: 05418, fold enriched = 2.61, FDR = 2.05×10^{-5}) (Figure 3B).

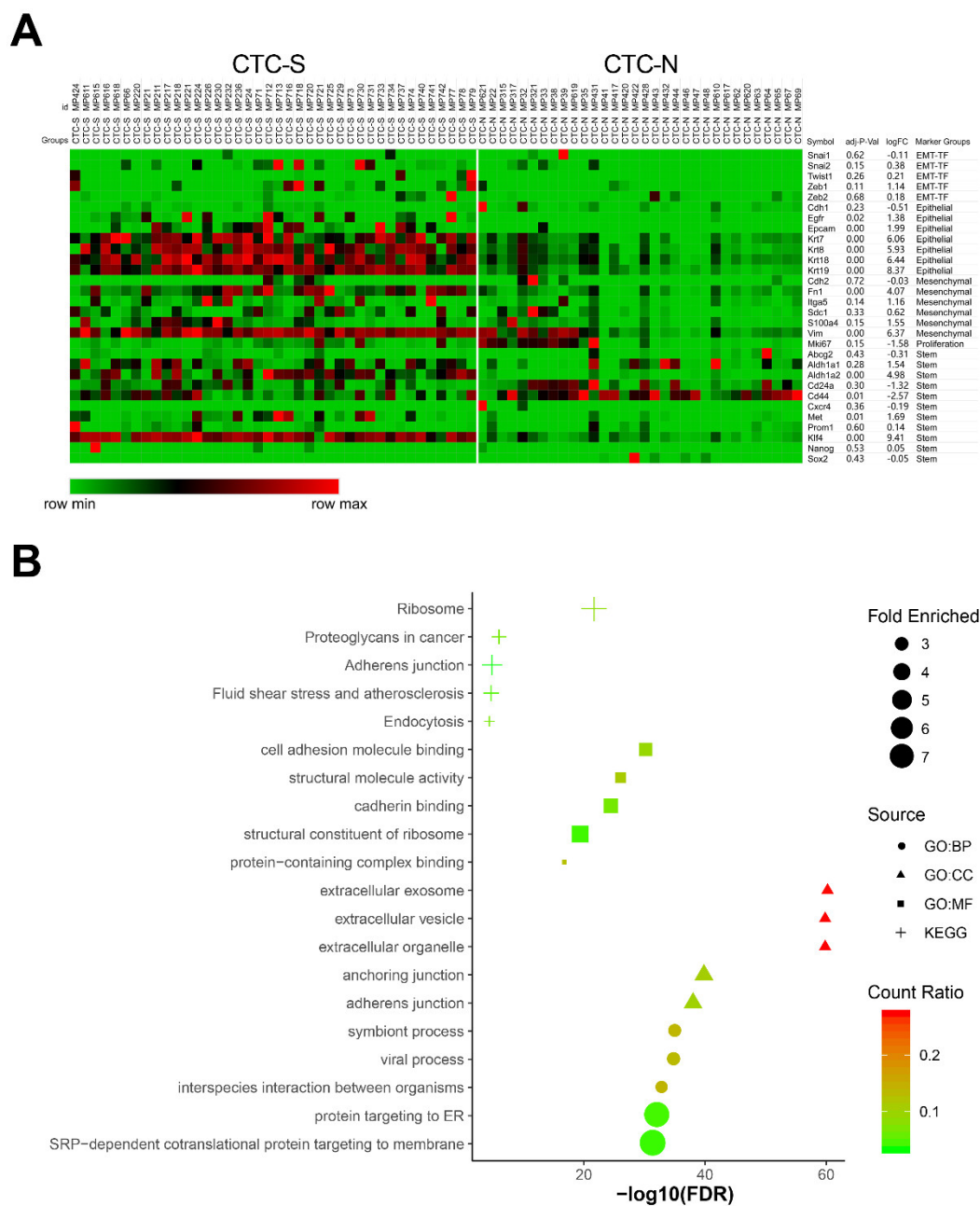


Figure 3. Differentially expressed genes between CTC-S and CTC-N groups. **(A)** MORPHEUS-generated heatmap showing the expression of the marker set in CTC-S and CTC-N groups. The maximum and the minimum values in each row are displayed red and green, respectively. The marker names, groups, $\log_2(\text{Fold Change (FC)})$ values, and false discovery rate (FDR) values were listed on the right side. **(B)** The functional enrichment analysis of up-regulated genes in the CTC-S subgroup was visualized by imageGP (available online: <http://www.ehbio.com/ImageGP/index.php/Home/Index/index.html>, 23 April 2020), top 5 enriched Kyoto Encyclopedia of Genes and Genomes (KEGG) pathways, Gene Ontology (GO) terms for biologic processes (BP), cell components (CC) and molecular functions (MF), each aspect was shown (fold enriched >2 and FDR <0.010).

In order to gain deeper insight into the molecular biology of the CTC subsets, we used a weighted gene co-expression network analysis (WGCNA) in this study.

After network construction (Figure S4A, B), we identified five gene co-expression modules (blue, brown, green, turquoise, and yellow) with sizes between 67 and 1384 genes in the 72 samples (Figure

4A). Both brown and turquoise modules correlated with stemness (correlation coefficient >0.7 , adjusted p -value <0.001), but the results of the brown module were more significant (correlation coefficient = 0.95, adjusted p -value = 3.6×10^{-37}).

We therefore used the brown module in the functional enrichment analysis. In the brown module, 645 genes were functionally enriched in the adherens junction pathway (KEGG: 04520, fold enriched = 3.71, FDR = 3.59×10^{-3}), which is consistent with the results of the analysis of the differentially expressed genes. In addition, anchoring junction (GO: 0070161, fold enriched = 3.27, FDR = 2.22×10^{-13}), adherens junction organization (GO: 0034332, fold enriched = 4.29, FDR = 8.11×10^{-3}) and regulation of adherens junction organization (GO: 1903391, fold enriched = 13.11, FDR = 8.12×10^{-3}) were all significantly enriched again (Figure 4B).

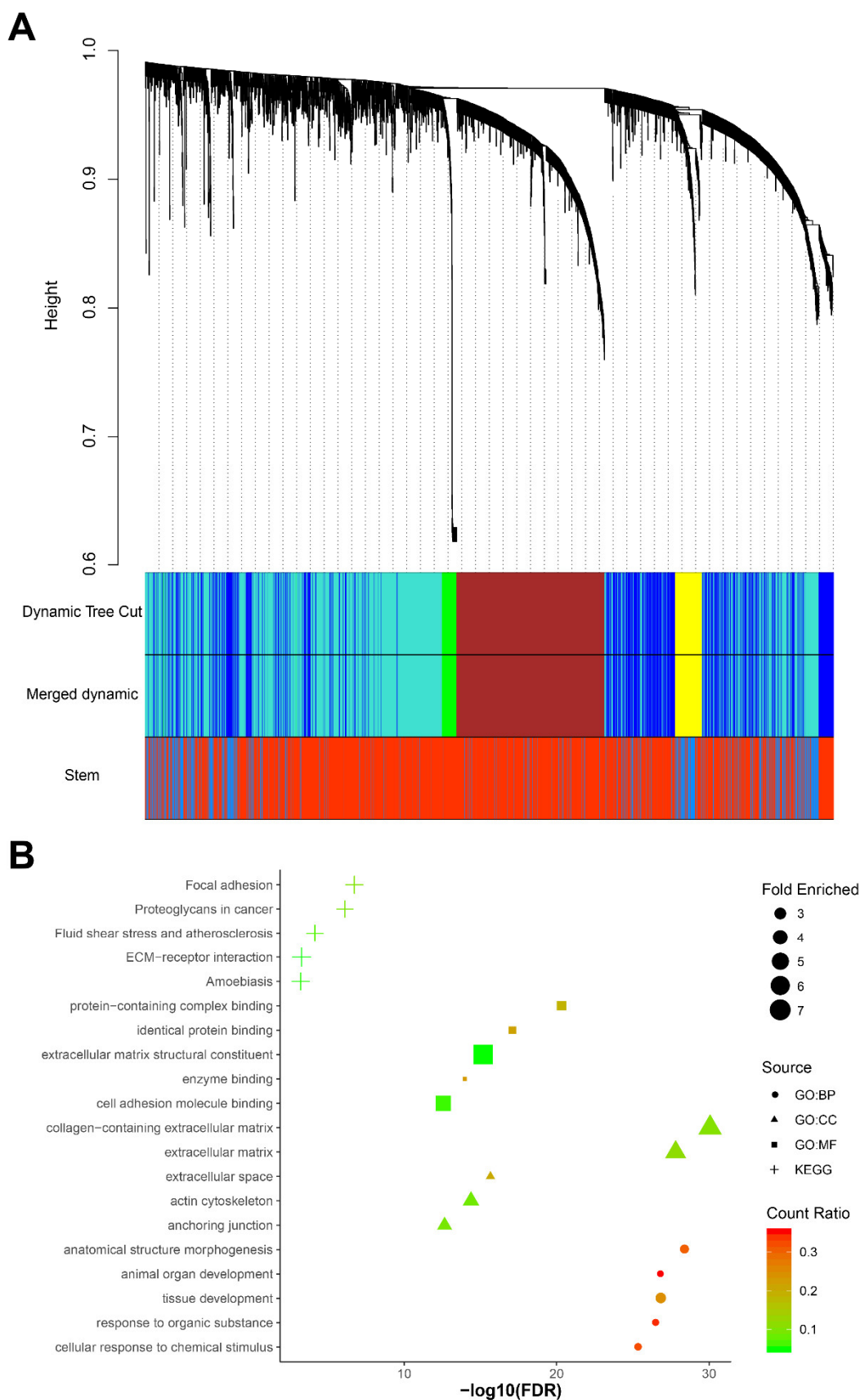


Figure 4. Weighted gene co-expression network analysis (WGCNA) of 72 pancreatic CTCs. **(A)** The cluster dendrogram of 3000 genes, five modules (blue, brown, green, turquoise, and yellow), were identified. The brown module correlates with the stem trait most significantly ($r = 0.95$, adjusted p -value = 3.6×10^{-3}). **(B)** The functional enrichment analysis of genes in the brown module (generated

by imageGP, available online: <http://www.ehbio.com/ImageGP/index.php/Home/Index/index.html>, accessed on 23 April 2020), top 5 enriched Kyoto Encyclopedia of Genes and Genomes (KEGG) pathways, Gene Ontology (GO) terms for biological processes (BP), cell components (CC) and molecular functions (MF) are shown [fold enriched > 2 and false discovery rate (FDR) <0.010].

To identify the pathways with the strongest influence on the CTC-S phenotype, we intersected functional enrichment pathways from the results of differentially expressed genes and WGCNA. This resulted in six pathways, including focal adhesion (KEGG: 04510), adherens junction (KEGG: 04520), ECM-receptor interaction (KEGG: 04512), AGE-RAGE signaling pathway in diabetic complications (KEGG: 04933), proteoglycans in cancer (KEGG: 05205) and fluid shear stress and atherosclerosis (KEGG: 05418) (Figure 5A). Since adherens junction organization (GO: 0034332) was also significantly enriched in the GO: BP aspect of functional enrichment analysis of both differentially expressed genes and WGCNA, we identified the adherens junction as a pathway of interest in CTC-S. In addition, double enriched GO terms, such as the regulation of adherens junction organization (GO: 1903391), cell–cell junction (GO: 0005911), and cell–cell junction assembly (GO: 0007043) provide further evidence that adherens junctions are biologically relevant in the CTC-S group.

In order to identify the key genes in the adherens junction pathway and the entire PPI network, hub genes of up-regulated genes in the PPI, genes enriched in the adherens junction pathway of differentially expressed genes and the WGCNA were intersected. The gene *Ctnnb1* ($\log_2FC = 3.24$, $FDR = 2.40 \times 10^{-4}$) was found significant in all three analyses (Figure 5B). The adherens junction pathway map and gene expression heatmap are plotted in Figures 5C and S5.

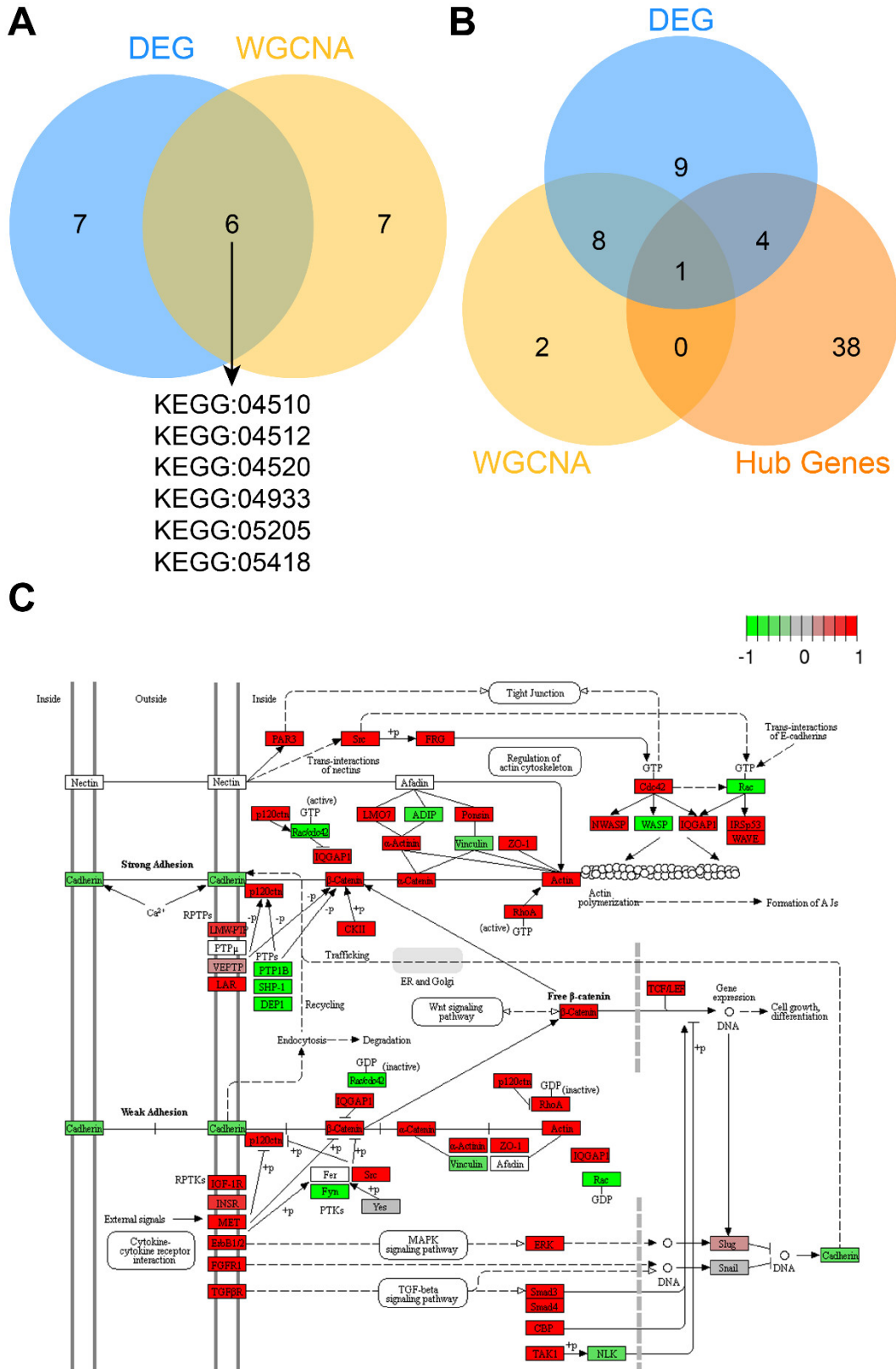


Figure 5. The adherens junctions pathway is significantly enriched in the CTC-S group. (A) Venn plot of enriched pathways in differentially expressed genes and weighted gene co-expression network analysis (WGCNA). Six pathways included focal adhesion (KEGG: 04510), ECM-receptor interaction (KEGG: 04512), adherens junction (KEGG: 04520), AGE-RAGE signaling pathway in diabetic complications (KEGG: 04933), proteoglycans in cancer (KEGG: 05205) and fluid shear stress and atherosclerosis (KEGG: 05418) were significantly enriched in both differentially expressed genes and WGCNA. (B) The intersection of enriched genes in the adherens junction pathway of differentially expressed

genes, enriched genes in the adherens junction pathway of the brown module in WGCNA, and hub genes. *Ctnnb1* is at the intersection of all three lists. (C) Gene expression in the adherens junction pathway was plotted by PATHVIEW [33]. Red represents up-regulated genes, while green represents down-regulated genes in the CTC-S group. The Venn plots in this study were generated by imageGP (available online: <http://www.ehbio.com/ImageGP/index.php/Home/Index/index.html>, accessed on 23 April 2020).

2.4. *Ctnnb1* and *Klf4* Positively Correlate

Next, we evaluated the correlation between *Ctnnb1* and stemness/pluripotency markers (*Aldh1a2*, *Met*, and *Klf4*), which were significantly up-regulated in the CTC-S group (Figure 6). Among these, we found that *Ctnnb1* overexpression most significantly correlates with *Klf4* upregulation ($r = 0.50$, $FDR = 4.99 \times 10^{-5}$).

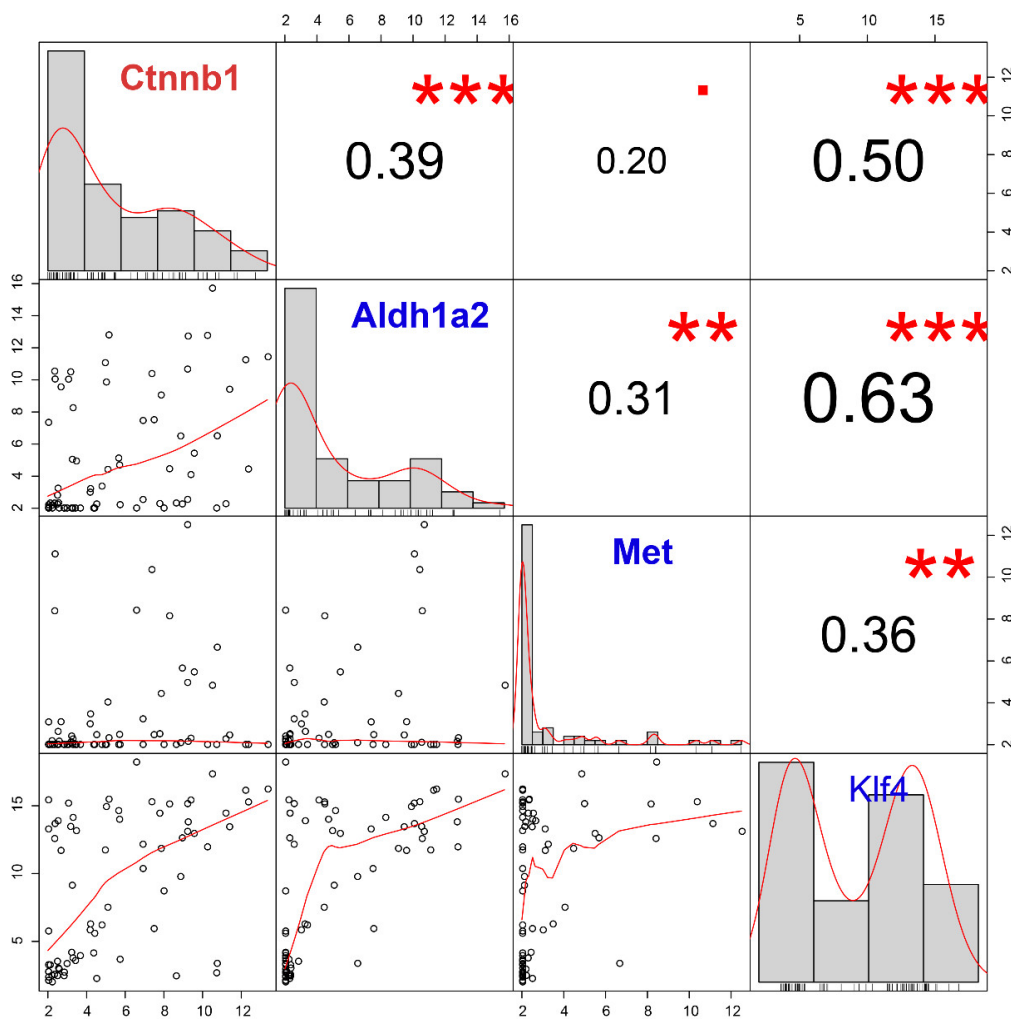


Figure 6. The correlation analysis of the candidate gene (*Ctnnb1*) and stem markers (*Aldh1a2*, *Met* and *Klf4*) demonstrates the positive correlation between *Klf4* and *Ctnnb1*. Gene expression histograms are represented on the diagonal. The lower left part is the scatter plots and the numbers in the upper right part correspond to the Pearson's correlation coefficient. *** $FDR < 0.001$; **, $0.001 < FDR < 0.010$; . $0.050 < FDR < 0.100$.

2.5. *CTNNB1* Is a Negative Prognostic Factor in Human PDAC

As reported above, *Klf4* and *Ctnnb1* were significantly up-regulated in murine pancreatic CTCs with stem-like features. To explore whether *KLF4* and *CTNNB1* are differentially expressed between pancreatic tumor and normal tissues, the visualization web tool Gene Expression Profiling Interactive Analysis 2 (GEPIA 2) (available online: <http://gepia2.cancer-pku.cn>, accessed on 7 March 2020) was

utilized [34]. When evaluating The Cancer Genome Atlas (TCGA) data, both *KLF4* and *CTNNB1* were up-regulated in PDAC compared to the matched normal tissues and the Genotype–Tissue Expression (GTEx) data [35] (Figure 7A,B).

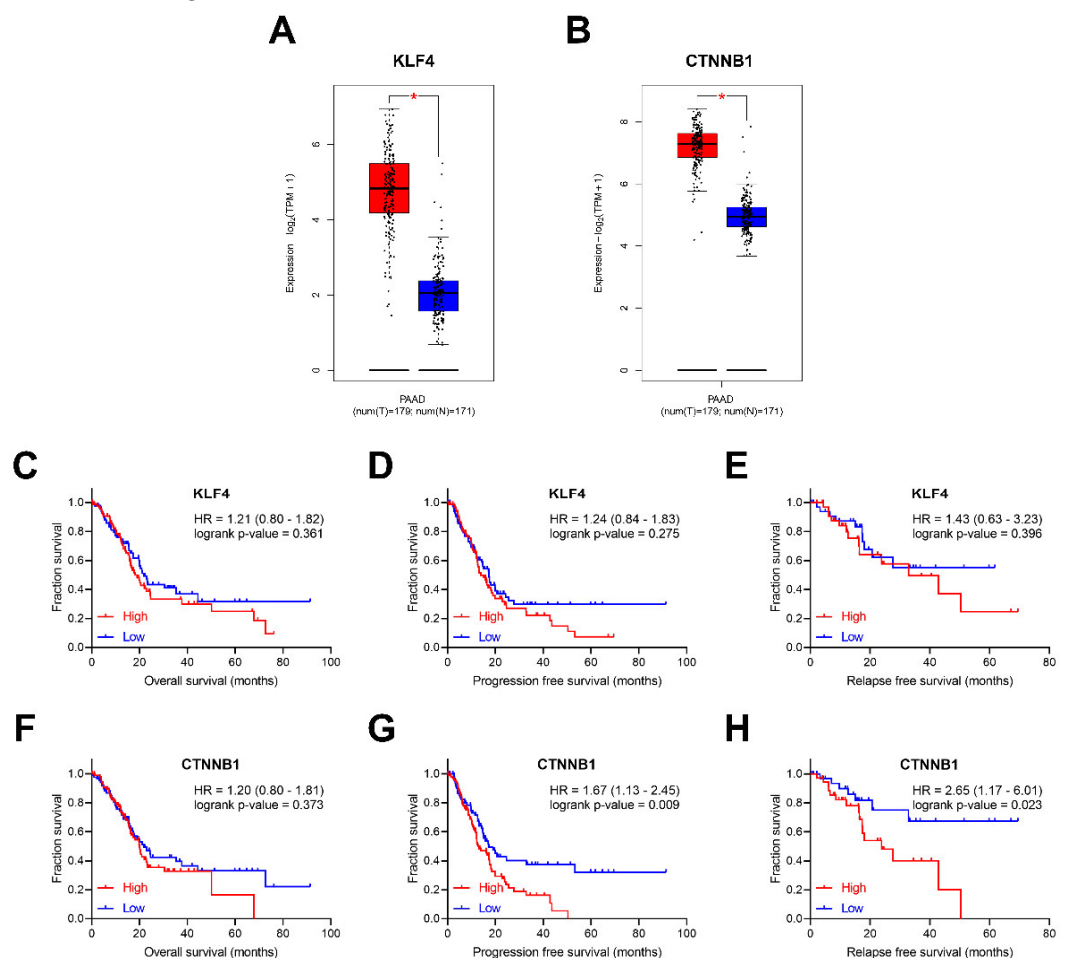


Figure 7. The clinical value of *KLF4* and *CTNNB1*. The box-plots with data overlaid dot plots for *KLF4* (A) and *CTNNB1* (B), showing expression in TCGA PAAD tumors with correspondent match PAAD normal tissue and GTEx pancreatic tissue. * p -value < 0.001. Clinical prognostic value of *KLF4*, overall survival (OS) (C), progression-free survival (PFS) (D), and relapse-free survival (RFS) (E). Clinical prognostic value of *CTNNB1*, OS (F), PFS (G) and RFS (H). Box plots were generated using GEPIA2 (available online: <http://gepia2.cancer-pku.cn/#index>, accessed on 7 March 2020).

We further investigated whether candidate genes expression levels correlate with clinical prognosis, including overall survival (OS), progression-free survival (PFS) and relapse-free survival (RFS) (Figure 7C–H) in the TCGA cohort [36]. No significant influence of *KLF4* on survival was observed. In contrast, although *CTNNB1* expression does not influence OS (Hazard Ratio (HR) = 1.20, log-rank p -value = 0.373), patients with higher *CTNNB1* expression have significantly worse PFS (HR = 1.67, log-rank p -value = 0.009) and RFS (HR = 2.65, log-rank p -value = 0.023).

3. Discussion

The aim of this study was to identify and characterize pancreatic CTCs with stem-like characteristics and explore potential underlying mechanisms through integrated bioinformatics analysis.

Numerous molecular markers, including *Cd24*, *Cd44*, *Prom1* (*Cd133*), and other genes, have been applied in order to define pancreatic cancer stem cells (CSCs) [30,37,38]. The CTC subgroup identified in this study as CTC-S demonstrated heterogeneous expression of stemness and pluripotency markers in comparison to CTC-N, with some markers showing increased expression (*Aldh1a2* and *Klf4*), some with reduced levels (*Abcg2*, *Cd44*, *Cxcr4*, *Nanog* and *Sox2*) and others with no significant change

in expression (*Aldh1a1*, *Cd24a*, *Met* and *Prom1*). The mutual exclusivity phenomenon may be due to the heterogeneous nature of stem-like cells and stemness markers [39]. Similar results have been published for breast cancer, where $CD44^{\text{high}} / CD24^{\text{low}}$ defines a cluster of tumor cells with stem-like features [40]. This mutual exclusivity is confirmed in the here-presented results.

The invasive capabilities (e.g., migration and invasion) of tumor cells are a prerequisite for successful initiation and completion of metastasis. Accumulating evidence suggests that these features can be acquired through epithelial–mesenchymal transition (EMT), which is considered as a hallmark of tumor cell dissemination [31,41]. The CTC-S subpopulation exhibits a mixed epithelial/mesenchymal phenotype, while the CTC-N group exhibits neither epithelial nor mesenchymal markers. This is consistent with previous studies relating EMT to stemness [42,43]. Furthermore, CTCs with partial EMT phenotype or stem-like features may predict unfavorable survival in cancers independently [44].

Epithelial markers (most prominently *EpCAM*) have been widely used as identification and isolation markers of CTCs, and the prognostic value of *EpCAM*-positive CTCs is evident [15,45–48], but previous data have pointed towards a relative down-regulation of epithelial markers on CTCs [13]. The here-presented results draw a more complex image of *EpCAM* expression in CTCs—it can be speculated that *EpCAM* expression on CTCs is a continuum from *EpCAM*-negative to highly *EpCAM*-positive cells. While, in this work, *EpCAM* seems to be relatively overexpressed on cells with presumably higher metastatic potential, more research on the role of *EpCAM* on CTCs is clearly required.

In theory, the loss of cell adhesion molecules during EMT should also coincide with the down-regulation of genes associated with adherens junctions. While both *Cdh1* and *Cdh2* (generally considered a mesenchymal marker) are downregulated in the CTC-S subgroup, most of the genes in the adherens junction pathway are up-regulated in comparison to CTC-N, resulting in a mixed epithelial/mesenchymal phenotype of CTC-S. This may represent a transitory state allowing CTC-S to undergo functional adaptation during the metastatic cascade [49]. Additionally, this overexpression of adherens junctions may enable CTC-S to bind to endothelial cells prior to extravasation.

Intercellular adhesion has been demonstrated to be crucial for successful metastasis of epithelial cancers [50]. Among the different types of intercellular adhesions, cell–cell adherens junctions are the most common and contribute to cell polarity, tissue architecture maintenance, cell movement limitation, and proliferation [50]. Strong adherens junctions are mediated by the cadherin–catenin complex [51–53]. The here-presented data show that the CTC-S cells express low levels of E-cadherin (*Cdh1*), while the other genes from the adherens junction pathway were up-regulated, such as *Rhoa* and *Cdc42*, both of which are necessary for adherens junction maintenance [54,55]. We hypothesized that CTC-S cells are in the process of forming the adherens junction through actin cytoskeleton remodeling [56].

Besides, another critical component of adherens junctions, plakoglobin (also known as γ -catenin), was found to be necessary for the formation of CTC clusters and further contributed to the metastatic cascade [57,58]. In summary, adherens junctions are critical to the formation of CTC clusters, which can form tumor-microemboli, ultimately outgrowing to overt metastases [57]. These cellular aggregates have been detected in 81% of PDAC patients with unfavorable OS and PFS [59]. However, it must be taken into account that all 75 pancreatic CTCs analyzed in this study were isolated individually, which means there are no data derived from CTC clusters in this dataset; the validity of the here-presented data concerning CTC clusters is therefore limited.

Numerous studies have explored the *Wnt* signaling pathway, mainly due to the fact that canonical *Wnt* signaling regulates, stabilizes, and promotes the accumulation of the β -catenin by inhibiting its degradation [60,61]. A few studies have demonstrated that *Wnt*/ β -catenin signaling plays a crucial role in the plasticity of stem cells [62,63]. Furthermore, *Wnt* signaling is activated in pancreatic CTCs according to the original study of GSE51372 [64]. Unexpectedly, we found no significant correlation between *Wnt* family members and *Ctnnb1* in the pancreatic CTC-S group. In fact, according to our results, these *Wnt* family members are even expressed at low levels (Figure S6). Given that *Klf4* is overexpressed in the CTC-S group, we believe that *Klf4* inhibits *Wnt* signaling in this context [65].

Besides, the β -catenin has been determined to be necessary for cell adhesion and pluripotency, even without Wnt signaling [66].

Klf4 (Krüppel-Like Factor 4) is enriched in CTC-S, which positively correlates with *Cttnb1*. As reported, *Klf4* is a versatile marker that can both suppress [67] or support [68] PDAC. Several studies have investigated *Klf4*, which encodes a protein that belongs to the Krüppel family of transcription factors and plays a vital role in maintaining embryonic or induced pluripotent stem cells as well as in preventing their differentiation [28,69,70]. In a recent study, Zheng et al. [71] found that *Klf4* promotes CTC survival by increasing intracellular reactive oxygen species. Intriguingly, *Klf4* can interact with *Cttnb1* through transcription regulation or direct protein interaction. As Tiwari et al. [72] reported, *Cttnb1* is one of the direct repression targets of the transcription factor *Klf4*. On the opposite side, β -catenin could also bind with the promoter of *Klf4* through activating the canonical Wnt pathway, as reported [73]. However, we found that the canonical Wnt family members are expressed at low levels, as previously mentioned. The evidence suggests that the interaction between *Klf4* and β -catenin might be happening through the formation of a protein complex rather than via the transcriptional regulation in the pancreatic CTC-S group. Even though there are studies claiming that *Klf4* inhibits β -catenin [74–76], there are other reports showing that the *Klf4* / β -catenin complex is necessary for the self-renewal capacity of stem/cancer cells [68]. The consensus is that the protein–protein interaction between *Klf4* and β -catenin happens in the nucleus [68,74–76]. According to previously published results [76,77], the cytoplasmic β -catenin could migrate to the nucleus freely, and there promote the transcription and translation of β -catenin targeted genes in conjunction with transcription factors. Among the β -catenin binding proteins extracted from epithelial cells nuclei, a 55-kDa protein could be identified as being *KLF4* [76]. Furthermore, the de novo synthesis of both E-cadherin and β -catenin increased at a similar rate in order to reconstruct an adherens junction after the proteolytic disruption of this extracellular interaction in epithelial cancer cells [77].

Clinically, *KLF4* expression in the primary tumor does significantly influence survival. *CTNNB1* is a negative prognostic marker for RFS and PFS. For OS, a trend can be seen, but it fails to reach statistical significance. This supports the hypothesis that the high expression of *Cttnb1* in CTC-S contributes to the formation of adherens junctions, which in turn may promote the survival of CTC-S in circulation [78]. However, it must be taken into account that the clinical validation was performed using RNA-seq expression data of bulk tumor cells from the primary tumor rather than single-cell RNA sequencing of CTCs, which limits its validity.

It is important to reinforce that this study is an in silico analysis of a single pre-existing dataset from a single pancreatic cancer model, since no other high-throughput sequencing datasets of pancreatic CTCs are publically available. To reduce the bias from limited study resources, we combined the analysis of differentially expressed genes, WGCNA, correlation analysis, and added clinical survival data. The limited sample size, and the absence of prognostic data directly associated with CTC expression data, may partially limit the results. With all that under consideration, more preclinical and clinical studies are necessary to confirm our findings. However, despite its exploratory nature, this study offers new insight into pancreatic CTCs with stem-like characteristics.

4. Materials and Methods

4.1. Data Collection, Data Quality Check, and Normalization

Single-cell RNA-sequencing data of 75 murine pancreatic CTCs with sufficient quality in dataset GSE51372 were downloaded from the Gene Expression Omnibus (GEO) database (available online: <http://www.ncbi.nlm.nih.gov/geo/>, accessed on 19 August 2019). Gene expression and clinical information profiles for Pancreas Adenocarcinoma (PAAD) patients were obtained from The Cancer Genome Atlas (TCGA) data portal (available online: <https://tcga-data.nci.nih.gov/tcga/>, accessed on 19 August 2019) [36].

In the pre-process part, an integrated web tool iDEP 9.0 (available online: <http://bioinformatics.sdstate.edu/idep/>, accessed on 7 March 2020) was employed to analyze the 75 CTCs sequencing data [79]. Three samples with a sequencing depth of less than two counts per million (CPM) were

excluded, genes with less than 0.5 CPM in all the samples were also filtered out, and then transformed the remaining 72 samples with EdgeR: $\log_2(\text{CPM} + c)$, pseudo count $c = 4$ [79]. (Figure S1) Mitochondrial RNA (mtRNA) was not used for QC as mtRNA is not contained in the GSE51372 read counts file.

4.2. Principal Component Analysis (PCA)

The top 3000 (approximately equal to 25%) [80] most variable genes were identified by the iDEP 9.0. Then, we used an open-source web tool named ClustVis (available online: <https://biit.cs.ut.ee/clustvis/>, accessed on 7 March 2020) to perform the PCA and generate the PCA score plot [81]. Imputation was deemed unnecessary since the data were normalized in the aforementioned part. PCA loading data were downloaded from ClustVis and visualized using imageGP (available online: <http://www.ehbio.com/ImageGP/index.php/>, accessed on 23 April 2020). Next, the corresponding stem markers (PCA loadings) were used to define the distinct PCA score cluster with stem-like features.

4.3. Identification of Differentially Expressed Genes

After pre-processing, read counts of 11,931 genes were used to identify the differentially expressed genes. Data analysis was performed using the R package DESeq2 [82]. $|\text{Fold change (FC)}| > 2$ and a corrected p -value, false discovery rate (FDR) < 0.050 (Benjamini–Hochberg procedure) were set as cutoffs.

4.4. Weighted Gene Co-Expression Network Analysis (WGCNA)

We constructed a gene co-expression network of the 3000 most variable genes using the WGCNA package [83] in R x64 3.6.1 based on Euclidean distance. Before the topological overlap matrix construction, we estimated the soft threshold β . The details are described in Figure S4. To identify the module correlated with stem-like traits, we used the dynamic tree-cut algorithm (automatic single block method) to cluster dendrogram branches into several modules and assigned them colors. The minimum module size is set at 30, and the modules with larger than 0.9 pairwise correlation were merged. Only the module which significantly positively correlated with stem-like traits was included in the subsequent study.

4.5. Definition of Hub Genes

The up-regulated genes were submitted to the STRING 11.0 database [32], and the Protein–Protein Interaction (PPI) network was reconstructed via the Cytoscape software, version 3.5.1 or higher [84]. In addition to text mining, other basic settings were selected, such as co-expression, co-occurrence, databases, experiments, gene fusion, and neighborhood as active interaction sources. Besides that, only interaction pairs that scored >0.9 in the network were selected.

We used the cytoHubba plugin in Cytoscape, which provides 12 topological analysis methods including cluster coefficient, degree, the density of maximum neighborhood component, edge percolated component, maximal clique centrality, maximum neighborhood component and six other centralities (betweenness, bottleneck, closeness, eccentricity, radiality, and stress) for ranking nodes [85]. If the analyzed genes ranked in the top 10% in more than eight of the aforementioned methods, they were defined as hub genes.

4.6. Functional Enrichment Analysis

We used g:Profiler (available online: <https://biit.cs.ut.ee/gprofiler/gost>, accessed on 23 March 2020) [86] to perform functional enrichment analysis. Several enriched pathways of the Kyoto Encyclopedia of Genes and Genomes (KEGG) [87] and functional interpretations of gene ontology (GO) [88], including biological processes (BP), cellular components (CC) and molecular functions (MF), were identified [28]. The enrichment of fold >2.0 and FDR <0.010 were set as cutoffs.

4.7. Kaplan–Meier Survival Plot

Pathologically confirmed PDAC samples and their corresponding RNA-seq data from TCGA were included in this experiment [20]. Pre-processed level 3 data generated using Illumina HiSeq 2000 RNA Sequencing V2 and gene symbols *KLF4* and *CTNNB1* were used. For each tumor sample of the patient, the expression level was determined using a combination of MapSplice and RSEM. The individual sample files were merged in R using the *plyr* package [89]. In the next visualization step, GraphPad Prism 8.2.1 was utilized for the Log-rank (Mantel–Cox) test and to generate the Kaplan–Meier plots.

4.8. Statistical Analysis

Correlations of gene expression were evaluated with *cor* function (Pearson method by default) in R with the *PerformanceAnalytics* package. We calculated the Benjamini–Hochberg FDR. Only FDR <0.050 was considered statistically significant.

Additional tools used in this study are listed in Table S1.

5. Conclusions

In summary, we performed an integrated bioinformatic analysis of single-cell RNA sequencing data of pancreatic CTCs derived from the GEMM. We identified two distinct cell populations with (CTC-S) and without (CTC-N) stem cell-like properties. Various markers, including EMT-transcription factors, epithelial, mesenchymal, stemness, pluripotency, and proliferation markers, were used to characterize the CTC-S population. The adherens junction pathway was found to be significantly enriched CTC-S. This pathway may be activated by up-regulated *Ctnnb1* (β -catenin) through its conjunction with transcription factor *Klf4*, thus enabling CTC-S to survive in the bloodstream and promote distant metastasis.

To conclude, this study suggests that pancreatic CTCs with stem-like features might survive in the bloodstream and reach target organs due to evaluated *Ctnnb1* expression and the activation of intracellular adherens junctions pathway.

Supplementary Materials: The following are available online at www.mdpi.com/2075-4418/10/5/305/s1, Figure S1: Pre-processing the raw data of the pancreatic circulating tumor cells (CTCs); Figure S2: The classification of 72 samples; Figure S3: The differential expressed genes; Figure S4: The construction of the weighted gene co-expression network of 72 pancreatic CTCs; Figure S5: The heatmap of genes involved in adherens junction pathway; Figure S6: Correlation analysis of *Ctnnb1* and *Wnt* family members in pancreatic CTC-S group. Table S1: Online web tools utilized in this study.

Author Contributions: Conceptualization, L.Z.; Data curation, L.Z. and B.G.; Formal analysis, L.Z., B.G. and J.-S.J.; Funding acquisition, L.Z.; Investigation, L.Z., B.G. and J.-C.J.; Methodology, L.Z.; Project administration, S.S.; Validation, L.Z., B.H. and S.S.; Visualization, L.Z., B.G. and S.S.; Writing—original draft, L.Z., B.H. and S.S.; Writing—review and editing, L.Z., B.H., B.G., J.-C.J., C.Y., C.R. and S.S. All authors have read and agreed to the published version of the manuscript.

Funding: L.Zhu is supported by the China Scholarship Council (CSC) for his doctoral degree, 201908080072.

Acknowledgments: L. Zhu would like to thank Xijin Ge, Department of Mathematics and Statistics, South Dakota State University, USA, for technical support on the iDEP.

Conflicts of Interest: The authors declare no conflict of interest. The funders had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, or in the decision to publish the results.

References

1. Siegel, R.L.; Miller, K.D.; Jemal, A. Cancer statistics, 2019. *CA Cancer J. Clin.* **2019**, *69*, 7–34.
2. Strobel, O.; Neoptolemos, J.; Jäger, D.; Büchler, M.W. Optimizing the outcomes of pancreatic cancer surgery. *Nat. Rev. Clin. Oncol.* **2019**, *16*, 11–26.
3. Ding, Y.; Cravero, J.D.; Adrian, K.; Grippo, P. Modeling Pancreatic Cancer In Vivo, From Xenograft and Carcinogen-Induced Systems to Genetically Engineered Mice. *Pancreas* **2010**, *39*, 283–292.
4. Eferl, R.; Casanova, E. *Mouse Models of Cancer: Methods and Protocols*; Humana Press: New York, NY, USA, 2015; Volume 1267, ISBN 978-1-4939-2297-0.
5. Aguirre, A.J.; Bardeesy, N.; Sinha, M.; Lopez, L.; Tuveson, D.A.; Horner, J.; Redston, M.S.; Depinho, R.A. Activated Kras and Ink4a/Arf deficiency cooperate to produce metastatic pancreatic ductal adenocarcinoma. *Genes Dev.* **2003**, *17*, 3112–3126.
6. Hingorani, S.R.; Wang, L.; Multani, A.S.; Combs, C.; Deramaudt, T.B.; Hruban, R.H.; Rustgi, A.K.; Chang, S.; Tuveson, D.A. Trp53R172H and KrasG12D cooperate to promote chromosomal instability and widely metastatic pancreatic ductal adenocarcinoma in mice. *Cancer Cell* **2005**, *7*, 469–483.
7. Massagué, J.; Obenauf, A.C. Metastatic colonization by circulating tumour cells. *Nature* **2016**, *529*, 298–306.
8. Court, C.M.; Ankeny, J.S.; Sho, S.; Winograd, P.; Hou, S.; Song, M.; Wainberg, Z.A.; Girgis, M.D.; Graeber, T.G.; Agopian, V.G.; et al. Circulating Tumor Cells Predict Occult Metastatic Disease and Prognosis in Pancreatic Cancer. *Ann. Surg. Oncol.* **2018**, *25*, 1000–1008.
9. Steinert, G.; Schölch, S.; Koch, M.; Weitz, J. Biology and significance of circulating and disseminated tumour cells in colorectal cancer. *Langenbecks Arch. Surg.* **2012**, *397*, 535–542.
10. Rahbari, N.N.; Bork, U.; Kircher, A.; Nimitz, T.; Schölch, S.; Kahlert, C.; Schmidt, T.; Steinert, G.; Ulrich, A.B.; Reissfelder, C.; et al. Compartmental differences of circulating tumor cells in colorectal cancer. *Ann. Surg. Oncol.* **2012**, *19*, 2195–2202.
11. Bork, U.; Grützmann, R.; Rahbari, N.N.; Schölch, S.; Distler, M.; Reissfelder, C.; Koch, M.; Weitz, J. Prognostic relevance of minimal residual disease in colorectal cancer. *World J. Gastroenterol* **2014**, *20*, 10296–10304.
12. Schölch, S.; Bork, U.; Rahbari, N.N.; García, S.; Swiersy, A.; Betzler, A.M.; Weitz, J.; Koch, M. Circulating tumor cells of colorectal cancer. *Cancer Cell Microenviron.* **2014**, *1*, e323.
13. Steinert, G.; Schölch, S.; Niemietz, T.; Iwata, N.; García, S.A.; Behrens, B.; Voigt, A.; Kloor, M.; Benner, A.; Bork, U.; et al. Immune Escape and Survival Mechanisms in Circulating Tumor Cells of Colorectal Cancer. *Cancer Res.* **2014**, *74*, 1694–1704.
14. van Noort, V.; Schölch, S.; Iskar, M.; Zeller, G.; Ostertag, K.; Schweitzer, C.; Werner, K.; Weitz, J.; Koch, M.; Bork, P. Novel Drug Candidates for the Treatment of Metastatic Colorectal Cancer through Global Inverse Gene-Expression Profiling. *Cancer Res.* **2014**, *74*, 5690–5699.
15. Bork, U.; Rahbari, N.N.; Schölch, S.; Reissfelder, C.; Kahlert, C.; Büchler, M.W.; Weitz, J.; Koch, M. Circulating tumour cells and outcome in non-metastatic colorectal cancer: A prospective study. *Br. J. Cancer* **2015**, *112*, 1306–1313.
16. Rahbari, N.N.; Bork, U.; Schölch, S.; Reissfelder, C.; Thorlund, K.; Betzler, A.; Kahlert, C.; Schneider, M.; Ulrich, A.B.; Büchler, M.W.; et al. Metastatic Spread Emerging From Liver Metastases of Colorectal Cancer: Does the Seed Leave the Soil Again? *Ann. Surg.* **2016**, *263*, 345–352.
17. Schölch, S.; García, S.A.; Iwata, N.; Niemietz, T.; Betzler, A.M.; Nanduri, L.K.; Bork, U.; Kahlert, C.; Thepkaysone, M.-L.; Swiersy, A.; et al. Circulating tumor cells exhibit stem cell characteristics in an orthotopic mouse model of colorectal cancer. *Oncotarget* **2016**, *7*, 27232–27242.
18. Nanduri, L.K.; Hissa, B.; Weitz, J.; Schölch, S.; Bork, U. The prognostic role of circulating tumor cells in colorectal cancer. *Expert Rev. Anticancer Ther.* **2019**, *19*, 1077–1088.
19. Baccelli, I.; Schneeweiss, A.; Riethdorf, S.; Stenzinger, A.; Schillert, A.; Vogel, V.; Klein, C.; Saini, M.; Bäuerle, T.; Wallwiener, M.; et al. Identification of a population of blood circulating tumor cells from breast cancer patients that initiates metastasis in a xenograft assay. *Nat. Biotechnol.* **2013**, *31*, 539–544.
20. Barriere, G.; Fici, P.; Gallerani, G.; Fabbri, F.; Zoli, W.; Rigaud, M. Circulating tumor cells and epithelial, mesenchymal and stemness markers: Characterization of cell subpopulations. *Ann. Transl. Med.* **2014**, *2*, 109.
21. Nagrath, S.; Jack, R.M.; Sahai, V.; Simeone, D.M. Opportunities and Challenges for Pancreatic Circulating Tumor Cells. *Gastroenterology* **2016**, *151*, 412–426.

22. Agnoletto, C.; Corrà, F.; Minotti, L.; Baldassari, F.; Crudele, F.; Cook, W.J.J.; Di Leva, G.; D'Adamo, A.P.; Gasparini, P.; Volinia, S. Heterogeneity in circulating tumor cells: The relevance of the stem-cell subset. *Cancers* **2019**, *11*, 483.
23. Ting, D.T.; Wittner, B.S.; Ligorio, M.; Jordan, N.V.; Shah, A.M.; Miyamoto, D.T.; Aceto, N.; Bersani, F.; Branigan, B.W.; Xega, K.; et al. Single-Cell RNA Sequencing Identifies Extracellular Matrix Gene Expression by Pancreatic Circulating Tumor Cells. *Cell Rep.* **2014**, *8*, 1905–1918.
24. Akrap, N.; Andersson, D.; Bom, E.; Gregersson, P.; Ståhlberg, A.; Landberg, G. Identification of Distinct Breast Cancer Stem Cell Populations Based on Single-Cell Analyses of Functionally Enriched Stem and Progenitor Pools. *Stem Cell Rep.* **2016**, *6*, 121–136.
25. Lapin, M.; Tjensvoll, K.; Oltedal, S.; Javle, M.; Smaaland, R.; Gilje, B.; Nordgård, O. Single-cell mRNA profiling reveals transcriptional heterogeneity among pancreatic circulating tumour cells. *BMC Cancer* **2017**, *17*, 1–10.
26. Shibue, T.; Weinberg, R.A. EMT, CSCs, and drug resistance: The mechanistic link and clinical implications. *Nat. Rev. Clin. Oncol.* **2017**, *14*, 611–629.
27. Singh, S.; Arcaroli, J.; Messersmith, W.; Vasiliou, V. Acetaldehyde and retinaldehyde-metabolizing enzymes in colon and pancreatic cancers. *Adv. Exp. Med. Biol.* **2015**, *815*, 281–294.
28. Yamane, M.; Ohtsuka, S.; Matsuura, K.; Nakamura, A.; Niwa, H. Overlapping functions of Krüppel-like factor family members: Targeting multiple transcription factors to maintain the naïve pluripotency of mouse embryonic stem cells. *Development* **2018**, *145*, 1–12.
29. Scholzen, T.; Gerdes, J. The Ki-67 Protein: From the Known and the Unknown. *J. Cell. Phys.* **2000**, *322*, 311–322.
30. Ishiwata, T.; Matsuda, Y.; Yoshimura, H.; Sasaki, N.; Ishiwata, S.; Ishikawa, N.; Takubo, K.; Arai, T.; Aida, J. Pancreatic cancer stem cells: Features and detection methods. *Pathol. Oncol. Res.* **2018**, *24*, 797–805.
31. Zhou, P.; Li, B.; Liu, F.; Zhang, M.; Wang, Q.; Liu, Y.; Yao, Y.; Li, D. The epithelial to mesenchymal transition (EMT) and cancer stem cells: Implication for treatment resistance in pancreatic cancer. *Mol. Cancer* **2017**, *16*, 1–11.
32. Szklarczyk, D.; Gable, A.L.; Lyon, D.; Junge, A.; Wyder, S.; Huerta-Cepas, J.; Simonovic, M.; Doncheva, N.T.; Morris, J.H.; Bork, P.; et al. STRING v11: Protein-protein association networks with increased coverage, supporting functional discovery in genome-wide experimental datasets. *Nucleic Acids Res.* **2019**, *47*, D607–D613.
33. Luo, W.; Brouwer, C. Pathview: An R/Bioconductor package for pathway-based data integration and visualization. *Bioinformatics* **2013**, *29*, 1830–1831.
34. Tang, Z.; Kang, B.; Li, C.; Chen, T.; Zhang, Z. GEPIA2: An enhanced web server for large-scale expression profiling and interactive analysis. *Nucleic Acids Res.* **2019**, *47*, W556–W560.
35. Lonsdale, J.; Thomas, J.; Salvatore, M.; Phillips, R.; Lo, E.; Shad, S.; Hasz, R.; Walters, G.; Garcia, F.; Young, N.; et al. GTEx Consortium The Genotype-Tissue Expression (GTEx) project. *Nat. Genet.* **2013**, *45*, 580–585.
36. Raphael, B.J.; Hruban, R.H.; Aguirre, A.J.; Moffitt, R.A.; Yeh, J.J.; Stewart, C.; Robertson, A.G.; Cherniack, A.D.; Gupta, M.; Getz, G.; et al. Cancer Genome Atlas Research Network Integrated Genomic Characterization of Pancreatic Ductal Adenocarcinoma. *Cancer Cell* **2017**, *32*, 185–203.e13.
37. Lytle, N.K.; Ferguson, L.P.; Rajbhandari, N.; Gilroy, K.; Fox, R.G.; Deshpande, A.; Schürch, C.M.; Hamilton, M.; Robertson, N.; Lin, W.; et al. A Multiscale Map of the Stem Cell State in Pancreatic Adenocarcinoma. *Cell* **2019**, *177*, 572–586.e22.
38. Varillas, J.I.; Zhang, J.; Chen, K.; Barnes, I.I.; Liu, C.; George, T.J.; Fan, Z.H. Microfluidic isolation of circulating tumor cells and cancer stem-like cells from patients with pancreatic ductal adenocarcinoma. *Theranostics* **2019**, *9*, 1417–1425.
39. Islam, F.; Gopalan, V.; Lam, A.K.-Y. *Cancer Stem Cells, Methods and Protocols*, 1st ed.; Papaccio, G., Desiderio, V., Eds.; Humana Press: New York, NY, USA, 2018; ISBN 978-1-4939-7400-9.
40. Al-Hajj, M.; Wicha, M.S.; Benito-Hernandez, A.; Morrison, S.J.; Clarke, M.F. Prospective identification of tumorigenic breast cancer cells. *Proc. Natl. Acad. Sci. USA* **2003**, *100*, 3983–3988.
41. Brabletz, T.; Hlubek, F.; Spaderna, S.; Schmalhofer, O.; Hiendlmeyer, E.; Jung, A.; Kirchner, T. Invasion and Metastasis in Colorectal Cancer: Epithelial-Mesenchymal Transition, Mesenchymal-Epithelial Transition, Stem Cells and β -Catenin. *CTO* **2005**, *179*, 56–65.

42. Mani, S.A.; Guo, W.; Liao, M.-J.; Eaton, E.N.; Ayyanan, A.; Zhou, A.Y.; Brooks, M.; Reinhard, F.; Zhang, C.C.; Shipitsin, M.; et al. The epithelial-mesenchymal transition generates cells with properties of stem cells. *Cell* **2008**, *133*, 704–715.
43. Morel, A.-P.; Lièvre, M.; Thomas, C.; Hinkal, G.; Ansieau, S.; Puisieux, A. Generation of breast cancer stem cells through epithelial-mesenchymal transition. *PLoS ONE* **2008**, *3*, e2888.
44. Papadaki, M.A.; Stoupis, G.; Theodoropoulos, P.A.; Mavroudis, D.; Georgoulas, V.; Agelaki, S. Circulating Tumor Cells with Stemness and Epithelial-to-Mesenchymal Transition Features Are Chemoresistant and Predictive of Poor Outcome in Metastatic Breast Cancer. *Mol. Cancer Ther.* **2019**, *18*, 437–447.
45. Rahbari, N.N.; Aigner, M.; Thorlund, K.; Mollberg, N.; Motschall, E.; Jensen, K.; Diener, M.K.; Büchler, M.W.; Koch, M.; Weitz, J. Meta-analysis shows that detection of circulating tumor cells indicates poor prognosis in patients with colorectal cancer. *Gastroenterology* **2010**, *138*, 1714–1726.
46. Cohen, S.J.; Alpaugh, R.K.; Gross, S.; O'Hara, S.M.; Smirnov, D.A.; Terstappen, L.W.M.M.; Allard, W.J.; Bilbee, M.; Cheng, J.D.; Hoffman, J.P.; et al. Isolation and characterization of circulating tumor cells in patients with metastatic colorectal cancer. *Clin. Colorectal Cancer* **2006**, *6*, 125–132.
47. Cohen, S.J.; Punt, C.J.A.; Iannotti, N.; Saidman, B.H.; Sabbath, K.D.; Gabrail, N.Y.; Picus, J.; Morse, M.A.; Mitchell, E.; Miller, M.C.; et al. Prognostic significance of circulating tumor cells in patients with metastatic colorectal cancer. *Ann. Oncol.* **2009**, *20*, 1223–1229.
48. Cohen, S.J.; Punt, C.J.A.; Iannotti, N.; Saidman, B.H.; Sabbath, K.D.; Gabrail, N.Y.; Picus, J.; Morse, M.; Mitchell, E.; Miller, M.C.; et al. Relationship of circulating tumor cells to tumor response, progression-free survival, and overall survival in patients with metastatic colorectal cancer. *J. Clin. Oncol.* **2008**, *26*, 3213–3221.
49. Tam, W.L.; Weinberg, R.A. The epigenetics of epithelial-mesenchymal plasticity in cancer. *Nat. Med.* **2013**, *19*, 1438–1449.
50. Conacci-Sorrell, M.; Zhurinsky, J.; Ben-Ze'ev, A. The cadherin-catenin adhesion system in signaling and cancer. *J. Clin. Investig.* **2002**, *109*, 987–991.
51. Hinck, L.; Näthke, I.S.; Papkoff, J.; Nelson, W.J. Beta-catenin: A common target for the regulation of cell adhesion by Wnt-1 and Src signaling pathways. *Trends Biochem. Sci.* **1994**, *19*, 538–542.
52. Yamada, S.; Pokutta, S.; Drees, F.; Weis, W.I.; Nelson, W.J. Deconstructing the Cadherin-Catenin-Actin Complex. *Cell* **2005**, *123*, 889–901.
53. Drees, F.; Pokutta, S.; Yamada, S.; Nelson, W.J.; Weis, W.I. α -Catenin Is a Molecular Switch that Binds E-Cadherin- β -Catenin and Regulates Actin-Filament Assembly. *Cell* **2005**, *123*, 903–915.
54. Sahai, E.; Marshall, C.J. ROCK and Dia have opposing effects on adherens junctions downstream of Rho. *Nat. Cell Biol.* **2002**, *4*, 408–415.
55. Harris, T.J.C.; Tepass, U. Adherens junctions: From molecules to morphogenesis. *Nat. Rev. Mol. Cell Biol.* **2010**, *11*, 502–514.
56. Menke, A.; Giehl, K. Regulation of adherens junctions by Rho GTPases and p120-catenin. *Arch. Biochem. Biophys.* **2012**, *524*, 48–55.
57. Aceto, N.; Bardia, A.; Miyamoto, D.T.; Donaldson, M.C.; Wittner, B.S.; Spencer, J.A.; Yu, M.; Pely, A.; Engstrom, A.; Zhu, H.; et al. Circulating tumor cell clusters are oligoclonal precursors of breast cancer metastasis. *Cell* **2014**, *158*, 1110–1122.
58. Kemler, R. From cadherins to catenins: Cytoplasmic protein interactions and regulation of cell adhesion. *Trends Genet.* **1993**, *9*, 317–321.
59. Chang, M.-C.; Chang, Y.-T.; Chen, J.-Y.; Jeng, Y.-M.; Yang, C.-Y.; Tien, Y.-W.; Yang, S.-H.; Chen, H.-L.; Liang, T.-Y.; Wang, C.-F.; et al. Clinical Significance of Circulating Tumor Microemboli as a Prognostic Marker in Patients with Pancreatic Ductal Adenocarcinoma. *Clin. Chem.* **2016**, *62*, 505–513.
60. Janda, C.Y.; Dang, L.T.; You, C.; Chang, J.; de Lau, W.; Zhong, Z.A.; Yan, K.S.; Marecic, O.; Siepe, D.; Li, X.; et al. Surrogate Wnt agonists that phenocopy canonical Wnt/ β -catenin signaling. *Nature* **2017**, *545*, 234–237.
61. Gerlach, J.P.; Emmink, B.L.; Nojima, H.; Kranenburg, O.; Maurice, M.M. Wnt signalling induces accumulation of phosphorylated β -catenin in two distinct cytosolic complexes. *Open Biology* **2014**, *4*, 1–11.
62. Woo, D.; Yu, M. Circulating tumor cells as “liquid biopsies” to understand cancer metastasis. *Transl. Res.* **2018**, *201*, 128–135.
63. Zhang, P.; Chang, W.-H.; Fong, B.; Gao, F.; Liu, C.; Al Alam, D.; Bellusci, S.; Lu, W. Regulation of induced pluripotent stem (iPS) cell induction by Wnt/ β -catenin signaling. *J. Biol. Chem.* **2014**, *289*, 9221–9232.

64. Yu, M.; David, T.T.; Stott, S.L.; Wittner, B.S.; Ozsolak, F.; Paul, S.; Ciciliano, J.C.; Smas, M.E.; Winokur, D.; Gilman, A.J.; et al. RNA sequencing of pancreatic circulating tumour cells implicates WNT signaling in metastasis. *Nature* **2012**, *487*, 510–513.
65. Evans, P.M.; Chen, X.; Zhang, W.; Liu, C. KLF4 interacts with beta-catenin/TCF4 and blocks p300/CBP recruitment by beta-catenin. *Mol. Cell. Biol.* **2010**, *30*, 372–381.
66. Rudloff, S.; Kemler, R. Differential requirements for β -catenin during mouse development. *Development* **2012**, *139*, 3711–3721.
67. Zammarchi, F.; Morelli, M.; Menicagli, M.; Di Cristofano, C.; Zavaglia, K.; Paolucci, A.; Campani, D.; Arentini, P.; Boggi, U.; Mosca, F.; et al. KLF4 is a novel candidate tumor suppressor gene in pancreatic ductal carcinoma. *Am. J. Pathol.* **2011**, *178*, 361–372.
68. Hoffmeyer, K.; Raggioli, A.; Rudloff, S.; Anton, R.; Hierholzer, A.; Del Valle, I.; Hein, K.; Vogt, R.; Kemler, R. Wnt/ β -catenin signaling regulates telomerase in stem cells and cancer cells. *Science* **2012**, *336*, 1549–1554.
69. Karagiannis, P.; Takahashi, K.; Saito, M.; Yoshida, Y.; Okita, K.; Watanabe, A.; Inoue, H.; Yamashita, J.K.; Todani, M.; Nakagawa, M.; et al. Induced Pluripotent Stem Cells and Their Use in Human Models of Disease and Development. *Phys. Rev.* **2018**, *99*, 79–114.
70. Chronis, C.; Fiziev, P.; Papp, B.; Butz, S.; Bonora, G.; Sabri, S.; Ernst, J.; Plath, K. Cooperative Binding of Transcription Factors Orchestrates Reprogramming. *Cell* **2017**, *168*, 442–459.e20.
71. Zheng, Y.; Miyamoto, D.T.; Wittner, B.S.; Sullivan, J.P.; Aceto, N.; Jordan, N.V.; Yu, M.; Karabacak, N.M.; Comaills, V.; Morris, R.; et al. Expression of β -globin by cancer cells promotes cell survival during blood-borne dissemination. *Nat. Commun.* **2017**, *8*, 1–12.
72. Tiwari, N.; Meyer-Schaller, N.; Arnold, P.; Antoniadis, H.; Pachkov, M.; van Nimwegen, E.; Christofori, G. Klf4 is a transcriptional regulator of genes critical for EMT, including Jnk1 (Mapk8). *PLoS ONE* **2013**, *8*, e57329.
73. Ai, Z.; Shao, J.; Wu, Y.; Yu, M.; Du, J.; Shi, X.; Shi, X.; Zhang, Y.; Guo, Z. CHIR99021 enhances Klf4 Expression through β -Catenin Signaling and miR-7a Regulation in J1 Mouse Embryonic Stem Cells. *PLoS ONE* **2016**, *11*, e0150936.
74. Parenti, S.; Montorsi, L.; Fantini, S.; Mammoli, F.; Gemelli, C.; Atene, C.G.; Losi, L.; Frassinetti, C.; Calabretta, B.; Tagliafico, E.; et al. KLF4 Mediates the Effect of 5-ASA on the β -Catenin Pathway in Colon Cancer Cells. *Cancer Prev. Res.* **2018**, *11*, 503–510.
75. Sellak, H.; Wu, S.; Lincoln, T.M. KLF4 and SOX9 transcription factors antagonize β -catenin and inhibit TCF activity in cancer cells. *Biochim. Biophys. Acta* **2012**, *1823*, 1666–1675.
76. Zhang, W.; Chen, X.; Kato, Y.; Evans, P.M.; Yuan, S.; Yang, J.; Rychahou, P.G.; Yang, V.W.; He, X.; Evers, B.M.; et al. Novel cross talk of Kruppel-like factor 4 and beta-catenin regulates normal intestinal homeostasis and tumor repression. *Mol. Cell. Biol.* **2006**, *26*, 2055–2064.
77. Klinke, D.J.; Horvath, N.; Cuppett, V.; Wu, Y.; Deng, W.; Kanj, R. Interlocked positive and negative feedback network motifs regulate β -catenin activity in the adherens junction pathway. *Mol. Biol. Cell* **2015**, *26*, 4135–4148.
78. Hofmann, C.; Obermeier, F.; Artinger, M.; Hausmann, M.; Falk, W.; Schoelmerich, J.; Rogler, G.; Grossmann, J. Cell-Cell Contacts Prevent Anoikis in Primary Human Colonic Epithelial Cells. *Gastroenterology* **2007**, *132*, 587–600.
79. Ge, S.X.; Son, E.W.; Yao, R. iDEP: An integrated web application for differential expression and pathway analysis of RNA-Seq data. *BMC Bioinformatics* **2018**, *19*, 1–24.
80. Blischak, J. PCA of Highly Variable Genes. Available online: <https://jdblischak.github.io/singlecell-qt1/pca-variable.html> (accessed on 6 March 2020).
81. Metsalu, T.; Vilo, J. ClustVis: A web tool for visualizing clustering of multivariate data using Principal Component Analysis and heatmap. *Nucleic Acids Res.* **2015**, *43*, W566–W570.
82. Love, M.I.; Huber, W.; Anders, S. Moderated estimation of fold change and dispersion for RNA-seq data with DESeq2. *Genome Biol.* **2014**, *15*, 550.
83. Langfelder, P.; Horvath, S. WGCNA: An R package for weighted correlation network analysis. *BMC Bioinform.* **2008**, *9*, 559.
84. Shannon, P.; Markiel, A.; Ozier, O.; Baliga, N.S.; Wang, J.T.; Ramage, D.; Amin, N.; Schwikowski, B.; Ideker, T. Cytoscape: A Software Environment for Integrated Models of Biomolecular Interaction Networks. *Genome Res.* **2003**, *13*, 2498–2504.

85. Chin1, C.-H.; Chen, S.-H.; Wu, H.-H.; Ho, C.-W.; Ko, M.-T.; Lin, C.-Y. From cytoHubba: Identifying hub objects and sub-networks from complex interactome. *BMC Syst. Biol.* **2014**, *8*, 1–7.
86. Raudvere, U.; Kolberg, L.; Kuzmin, I.; Arak, T.; Adler, P.; Peterson, H.; Vilo, J. g:Profiler: A web server for functional enrichment analysis and conversions of gene lists (2019 update). *Nucleic Acids Res.* **2019**, *47*, W191–W198.
87. Kanehisa, M. Toward understanding the origin and evolution of cellular organisms. *Protein Sci.* **2019**, *28*, 1947–1951.
88. Ashburner, M.; Ball, C.A.; Blake, J.A.; Botstein, D.; Butler, H.; Cherry, J.M.; Davis, A.P.; Dolinski, K.; Dwight, S.S.; Eppig, J.T.; et al. Gene ontology: Tool for the unification of biology. The Gene Ontology Consortium. *Nat. Genet.* **2000**, *25*, 25–29.
89. Wickham, H. The split-apply-combine strategy for data analysis. *J. Stat. Softw.* **2009**, *40*, 174473.



© 2020 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<http://creativecommons.org/licenses/by/4.0/>).